# SOME ISSUES RELATED TO THE ACCURACY AND INTEPRETATION OF PLATELET VIABILITY MEASUREMENTS BY RADIO LABELING STUDIES

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Results of data presented were obtained from studies conducted at American Red Cross Research Department, Mid Atlantic Region

### ISSUES RELATED TO THE ACCURACY AND INTERPRETATION OF VIABILITY MEASUREMENTS

#### **Donor Variability in % Recovery**

- Inaccurate Estimation of Blood Volume
- Splenic Uptake
- Periodic variability

#### **Labeling Method/Procedure**

- Representative population
- Platelet Damage/Aggregates
- Isotope binding characteristics
- Contaminating cells

#### Data processing and interpretation

- Data points to Include
- Mathematical Models
  - Fitness of data
  - Robust and meaningful parameters

### ISSUES RELATED TO THE ACCURACY AND INTERPRETATION OF VIABILITY MEASUREMENTS

#### Variability in % Recovery related to the Donor

• Inaccurate Estimation of Blood Volume by body surface area

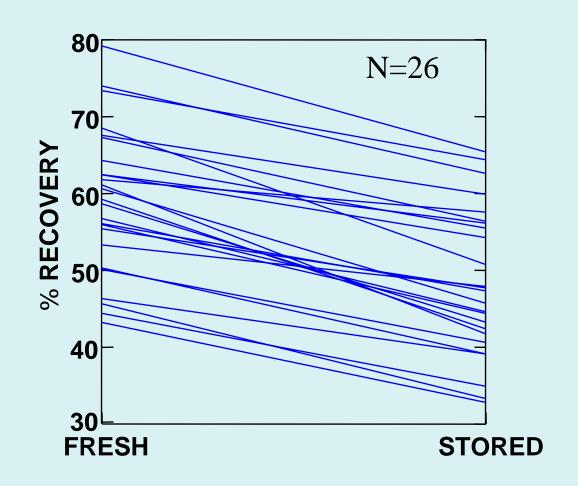
#### **Labeling Method**

Representative platelet population from the product

#### Data processing and interpretation

- Data points to include
- Mathematical Models
  - Fitness to raw data
  - Robust and meaningful parameters
  - Models comparing test to fresh platelets

## VARIABILITY IN % RECOVERY FRESH VS. 5 DAY STORED PC



#### VARIABILITY IN % RECOVERY

## SOURCE OF VARIABILITY WITH 5 DAY % RECOVERY BY REGRESSION ANALYSIS:

#### **SUM OF SQUARES:**

% OF TOTAL

Regression (fresh) 79 %

Residual (storage lesion) 21 %

(r squared = 0.79)

79 % of the variability is related to the recovery of fresh platelets from the donor and only 20 % is related to product platelet viability during 5 days storage

#### VARIABILITY IN % RECOVERY

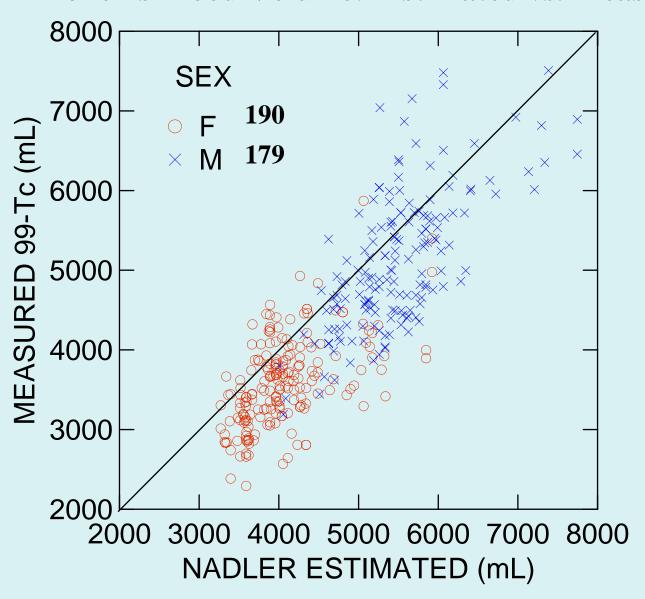
Importance of Accurate Estimation of Blood Volume:

% RECOVERY =

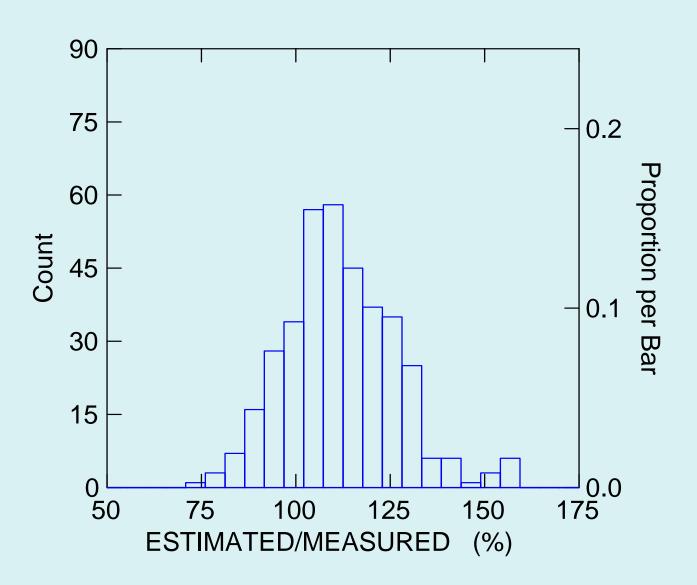
Radioactivity per mL Blood \* Blood Volume (mL) \* 100 % Radioactivity of the Injectate

Estimation of blood volume is commonly done by formulas (NADLER) for body surface area

### VARIABILITY IN % RECOVERY: Donor's Blood Volume: Estimated vs. Measured



#### VARIABILITY IN % RECOVERY: Blood Volume: Estimated vs. Measured



#### **VARIABILITY IN % RECOVERY**

A major source in variability in % recovery of a 5 day stored product is related to inaccurate estimation of the donor's blood volume and relatively little to the viability of the platelet product after storage:

- The determined % Recovery is not, by itself, an accurate measurement of the platelet viability of a 5 day standard product
- Paired Studies (test vs. control products from the same donor ) is thus preferable for determination of a potential change in platelet viability of a test as compared to a control product

## LABELING A REPRESENTATIVE PLATELET POPULATION OF THE TEST PRODUCT

#### Major assumption in radiolabeling studies:

Determination of platelet viability by radiolabeling is based on the assumption that platelets in the product population are uniformely labeled:

(that the amount of radioactivity per platelet is the same for all the platelets)

Thus, after infusion, a % decrease in radioactivity represents certain % loss of the number of injected platelets from circulation.

## LABELING A REPRESENTATIVE PLATELET POPULATION OF THE TEST PRODUCT

Assuming two populations of platelets in a product consisting of 50% viable and 50% damaged and nonviable platelets.

1)The uptake of isotope for the viable is 80% and for the non viable subpopulation 20% of total radioactivity.

After infusion the total population of the non viable is removed representing a loss of 50 % of the total platelet population. However, the loss of % radioactivity (% recovery) is only 20 %.

2) The non viable population are platelets that are lost during the labeling procedure.

Only the viable platelets are infused - no loss in % radioactivity (% recovery)

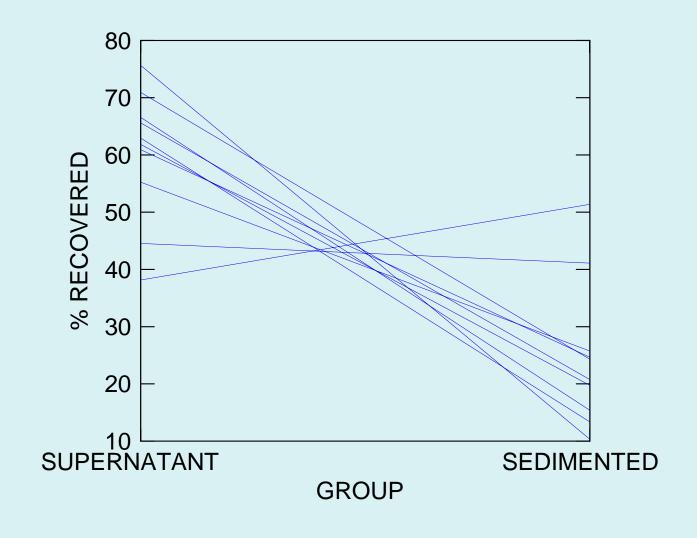
## LABELING A REPRESENTATIVE PLATELET POPULATION OF THE TEST PRODUCT

## Do platelet subpopulations from freshly collected whole blood differ in terms of viability?

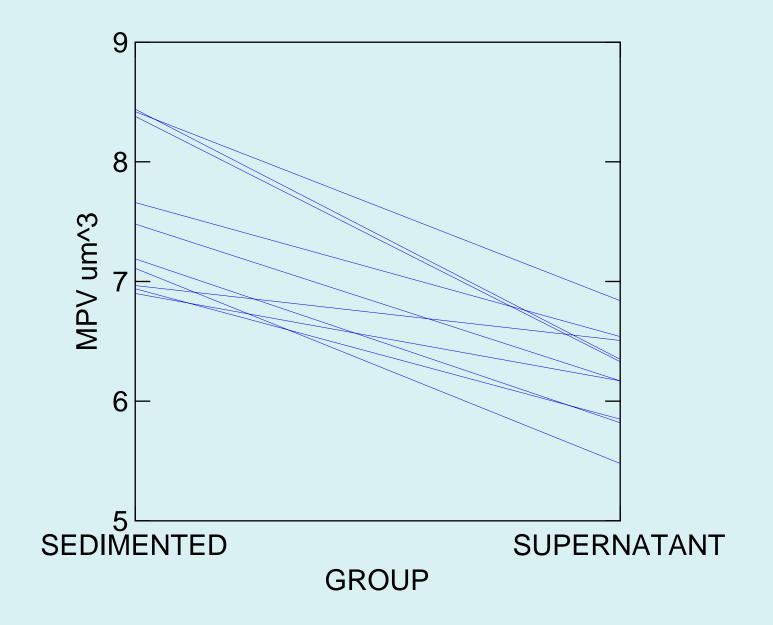
#### Study Design

- PRP (supernatant) platelet subpopulation was prepared by standard centrifugation using random donor WB units (n=8).
- The remaining buffy coat (sedimented) platelet subpopulation were obtained by additional processing.
- In vivo studies were conducted to determine viability of these two platelet subpopulations using simultaneous labeling and infusion with 111-In and 51-Cr

### LABELING OF PLATELET SUBPOPULATIONS: PLATELET COUNT RECOVERED FROM WHOLE BLOOD

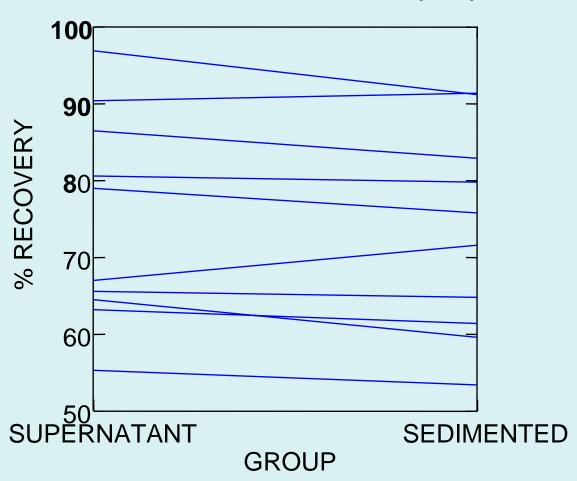


### LABELING OF PLATELET SUBPOPOULATIONS: PLATELET SIZE -MPV



### LABELING OF PLATELET SUBPOPULATIONS: IN VIVO VIABILITY - % RECOVERY

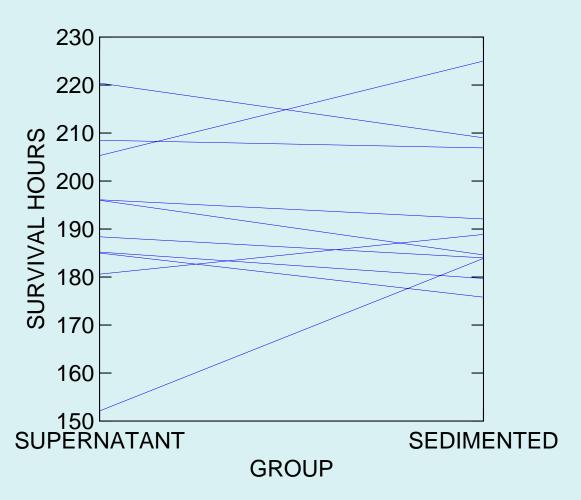
Mean SEDIMENTED = 73.2 (13.3) % Mean SUPERNATANT = 74.9 (13.7) %



#### LABELING OF PLATELET SUBPOPULATIONS:

SURVIVAL (NUMERIAL EXPECTED LIFESPAN)

Mean SEDIMENTED = 193 (17) HRS Mean SUPERNATANT = 192 (19) HRS



#### LABELING OF PLATELET SUB POPULATIONS

## Labeling of platelet subpopulations from freshly collected whole blood

#### Conclusions:

Using freshly collected blood two platelet subpopulations separated by size showed no statistically significant difference in % recovery and survival.

No statistically significant difference between results obtained using 111-In vs. 51-Cr .

#### DATA PROCESSING AND INTERPRETATION

#### Mathematical modeling of the raw data:

#### **Objective**

Reduce the data to a few accurate and meaningful parameters that be used to evaluate platelet viability of a product

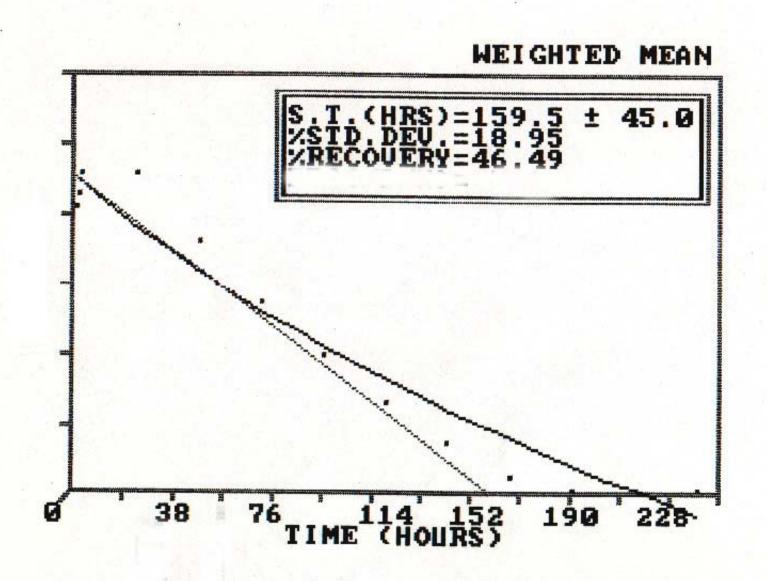
#### Data points to include?

#### **Method:**

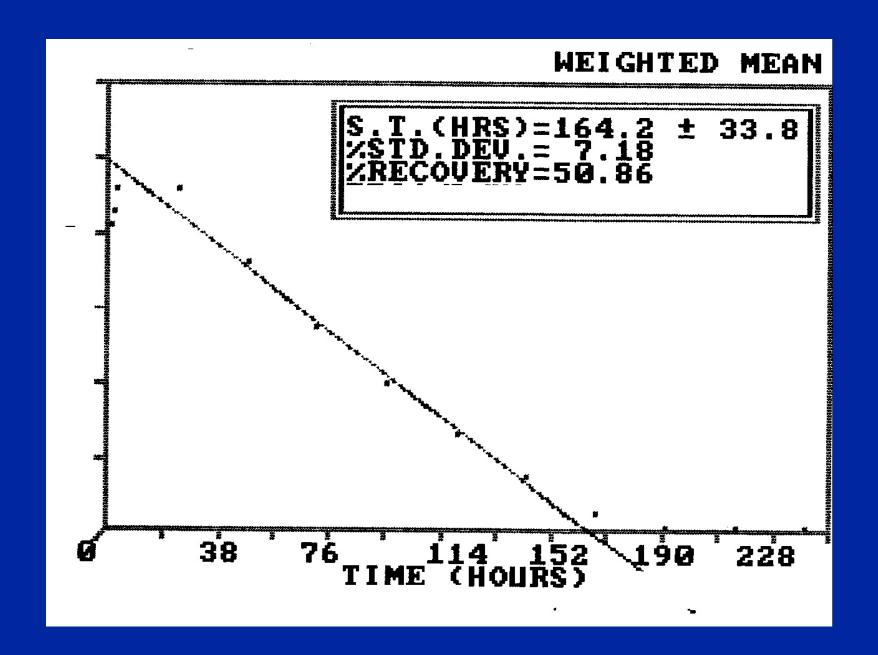
Least Sum Of Squares:

Minimize residual sum of squares = (Observed values – Model Predictions)^2 by iterative methods

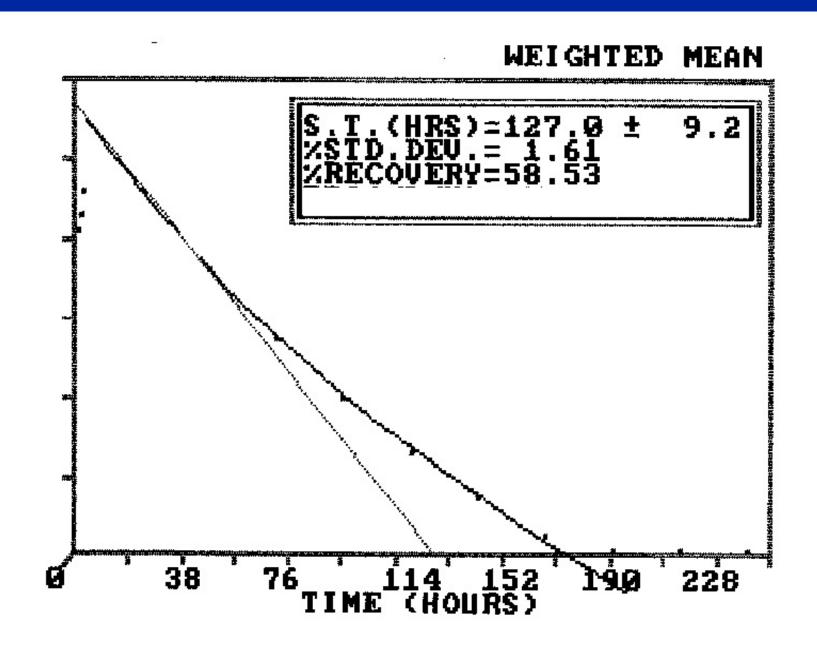
#### NUMERICAL EXPECTED LIFESPAN – All Data Points



#### NUMERICAL EXPECTED LIFESPAN – 3 h and daily Data Points



#### NUMERICAL EXPECTED LIFESPAN – 24 h and daily Data Points



#### MATHEMATICAL MODELING

#### Models used in platelet survivals:

Linear

**Exponential** 

**Multiple Hit (gamma function)** 

Weighted Mean

Meuleman

**Dornhorst** 

#### **Requirement:**

Must be able to fit a wide variety of typical survival curves for platelets stored/processed under various of conditions The goodness of fit is determined by the residual sum of squares

#### MEASUREMENTS OF PLATELET SURVIVAL

#### **Numerical Expected Lifespan:**

Intercept of the initial tangent of the survival curve with the x-axis (time)

#### Mean Residual Lifespan:

Area below the survival curve/%Recovery

#### T half:

Time after infusion at 50% of initial radioactivity

#### NUMERICAL EXPECTED LIFESPAN

#### **Definition:**

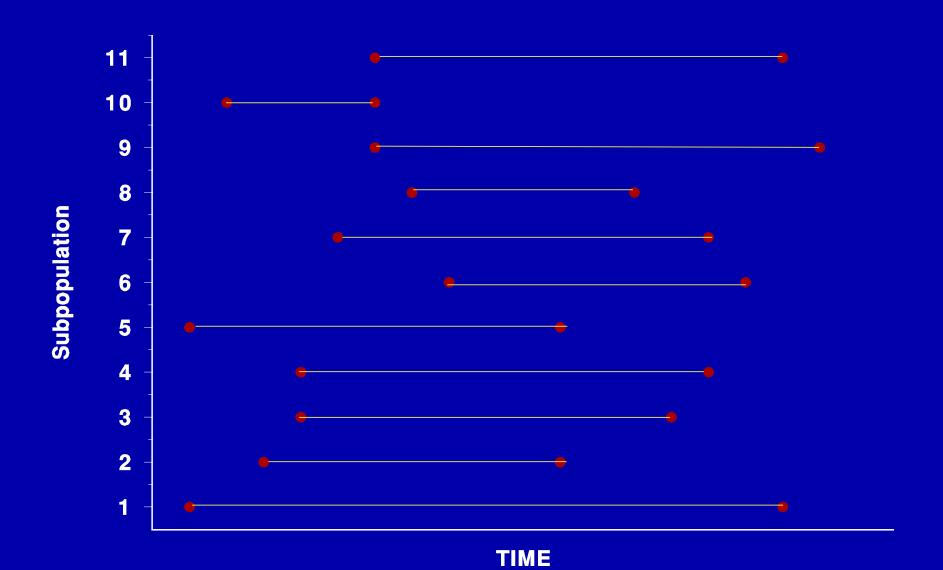
• Birth cohort lifespan of platelets newly released from the bone marrow

Used in estimation of platelet survivals in thrombocytopenic patients to determine:

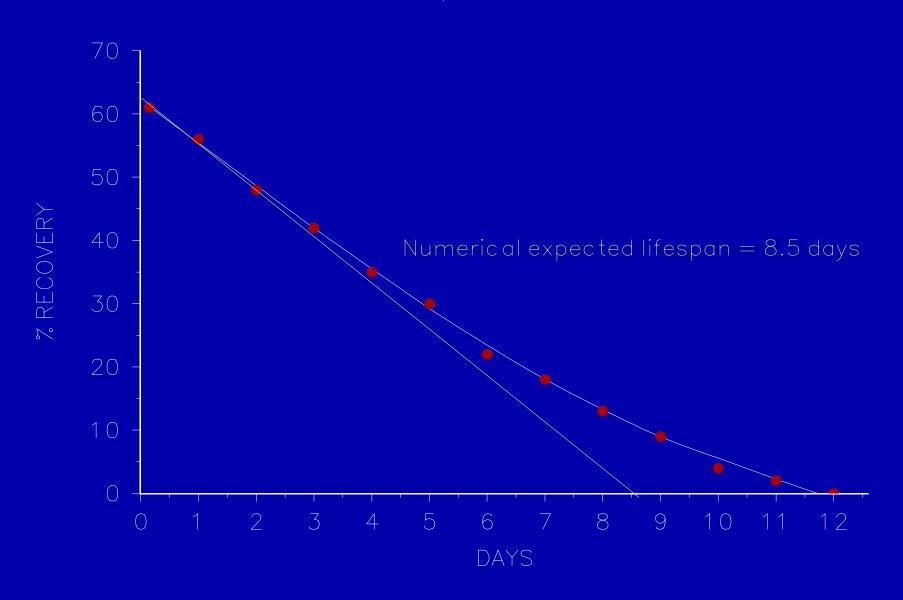
- Platelet turnover rates
- Events in the circulation system (senescence vs. random destruction

Meaningful in estimation of the survival of platelets in a product?

### NUMERICAL EXPECTED LIFESPAN 11 platelet subpopulations



### NUMERICAL EXPECTED LIFESPAN Fresh platelets

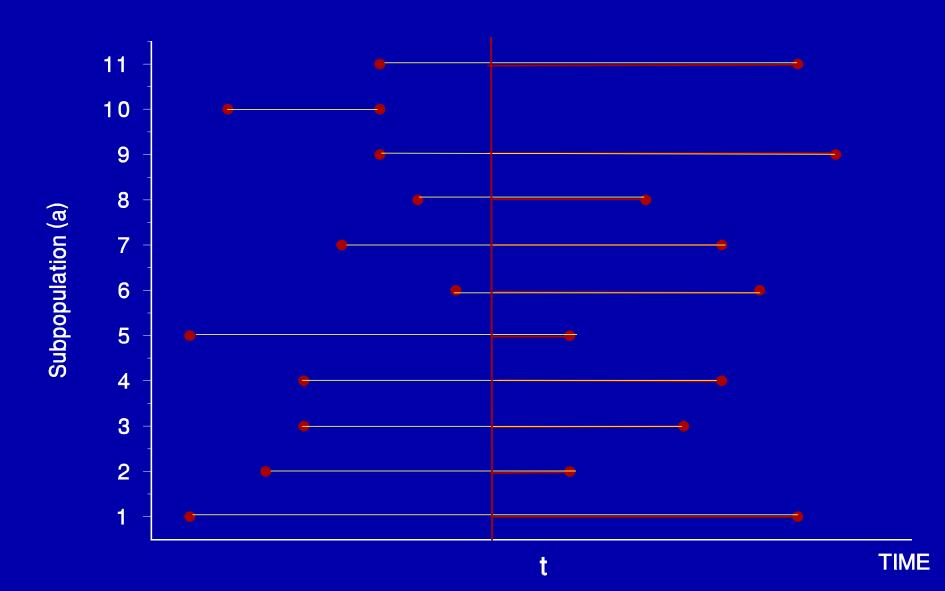


#### RESIDUAL LIFESPAN

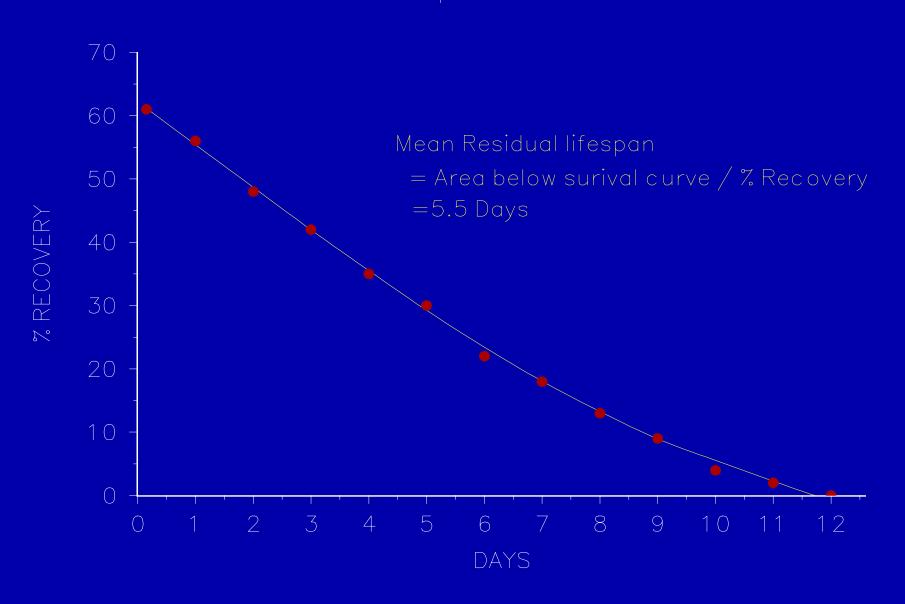
#### **Definition:**

- Mean residual lifespan in circulation of the labeled and infused platelet population (cross-sectional or sample population)
- More robust and meaningful in determination of the viability of a platelet product?

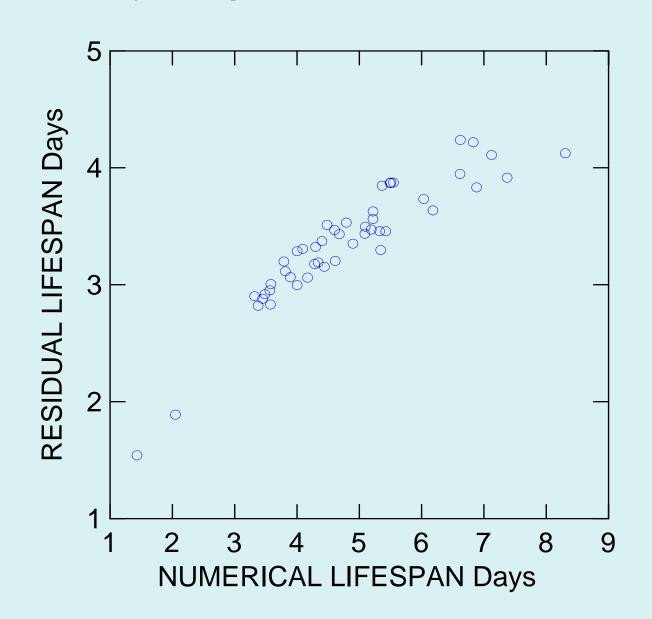
### CROSS SECTIONAL POPULATION 10 platelet subpopulations



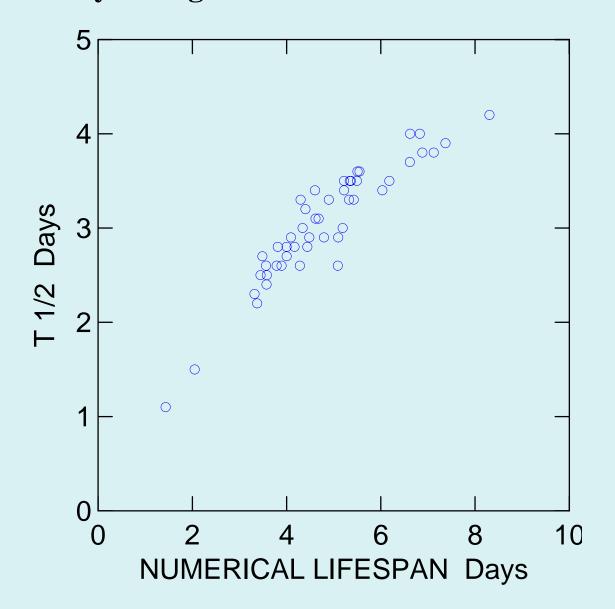
#### MEAN RESIDUAL LIFESPAN Fresh platelets



### RESIDUAL vs. NUMERICAL EXPECTED LIFESPAN (WMF) (5 vs. 7 day storage studies with RDPs in CLX bags)



T ½ vs. NUMERICAL EXPECTED LIFESPAN (WMF) (5 vs. 7 day storage studies with RDPs in CLX bags)



## SURVIVAL PARAMETERS: (5 vs 7 day storage studies with RDPs in CLX bags, Double label, n=24 pairs)

Parameter (by weighted mean function)	Day 5 mean	Day 7 mean	Difference 95 % CI paired t-test	Probability paired t-test
Numerical (Days)	5.3	4.4	0.5 – 1.4	<0.000
Residual (Days)	3.6	3.2	0.2 - 0.6	<0.000
T ½ (Days)	3.3	2.9	0.2 - 0.6	0.001
Random Destr. WMF (Exp. F.)	0.17	0.24	0.024- 0.12	0.005

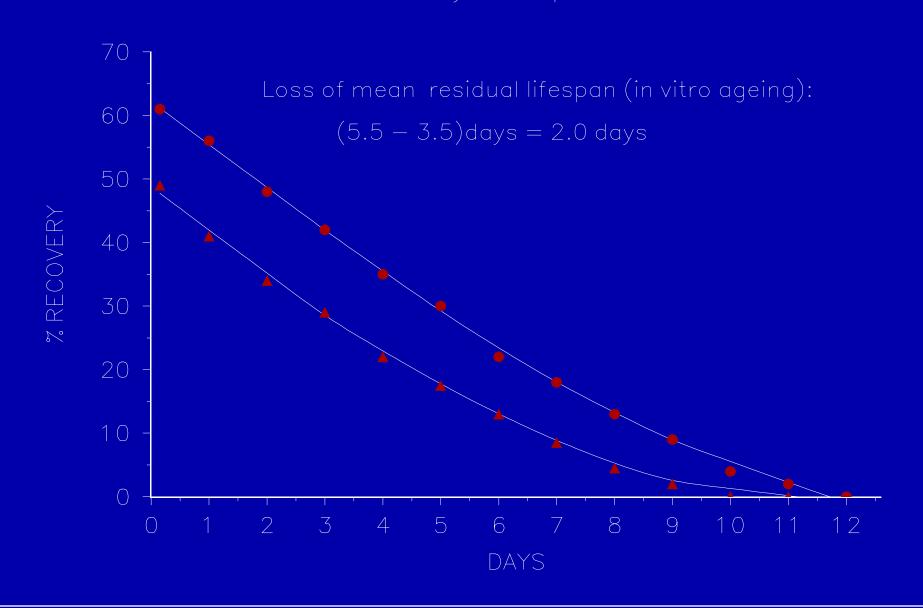
#### MATHEMATICAL MODELING

Comparison of the survival data of test platelets to that of fresh /control platelets may give useful information about the nature of a storage/process lesion.

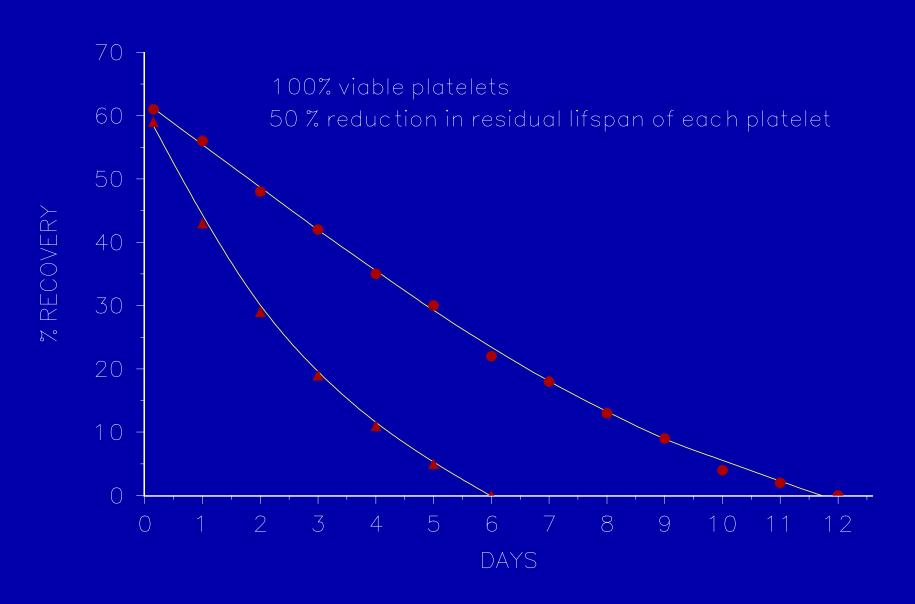
Some parameters that can be calculated by appropriate mathematical models are:

- Loss of % recovery due to aging versus due to random destruction
- Decrease in residual lifespan due to ageing versus random damage

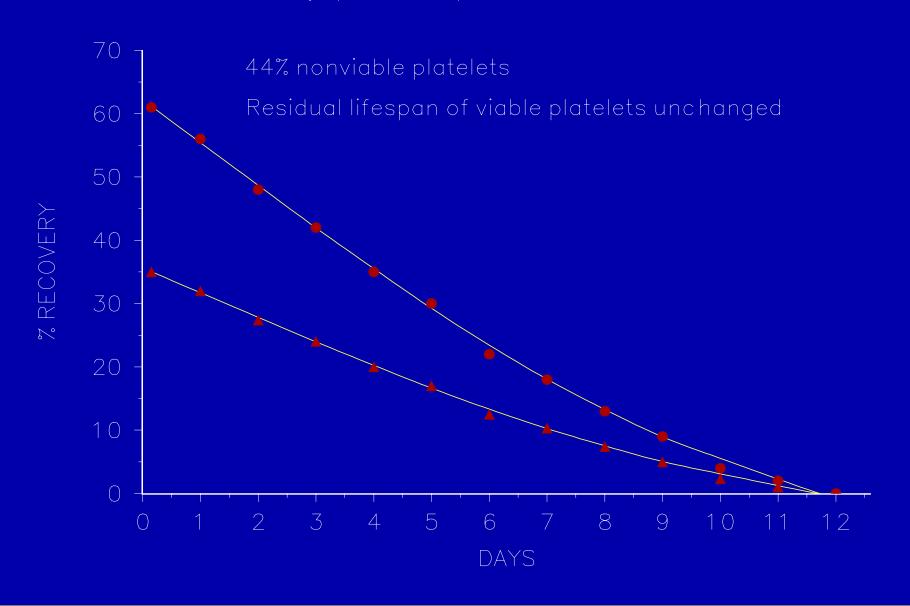
#### PLATELET IN VITRO AGEING Fresh and 5 day stored platelets



### PROPORTIONAL REDUCTION OF PLATELET LIFESPAN COLD EXPOSED PLATELETS



### RANDOM DESTRUCTION OF PLATELETS Cryopreserved platelets



### PROPOSED STEPS TO ENSURE ACCURACY OF VIABILITY MEASUREMENTS BY RADIO LABELING STUDIES

#### Variability Related To Donor

Inaccurate (and overestimated) Blood Volume based on current formulas for body surface area –

- Better formula for calculation of blood volume
- Paired studies

#### **Labeling Method**

Ensure uniform labeling of an representative population in a platelet product to be evaluated

- Determine platelet loss during labeling (test vs. control)
- Determine platelet size distribution pre and post labeling
- Determine Isotope uptake/elution in various subpopulations (test vs. control product)

### PROPOSED STEPS TO ENSURE APPROPRIATE DATA ANALYSIS AND INTERPRETATION

#### Select data points to be included based on

- Precision (more the better)
- Evenly spaced (clustered may cause biased results)
- Eliminate contribution of labeled RBCs

### Select appropriate mathematical models and parameters based on

- Goodness of fit by residual sum of squares
- Robustness
- Informative about the nature of a potential lesion/improvement of a product